Variations of the Nature of the Chiral Auxiliary with a Highly Enantioselective Chiral NADH Model

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Abstract: Various chiral amino alcohols **have been** used as chiral auxiliaries for a highly enantioselective NADH model. Some of these are new reagents which have been obtained by an enzymic resolution method.

Chiral NADH models are promising reagents in asymmetric reductions. In 1975, Ohno reported the first stereoselective reduction of ethyl benzoylformate by a 1,4-dihydronicotinamide derivative bearing a chiral carboxamide at C-3 (1). The corresponding (R)-mandelate was obtained with an enantiomeric excess of 20 %. Since this initial report, a large number of results have been described concerning the obtaining of high e.e. with performant chiral NADH models (2).

In all cases the reductions were performed in the presence of magnesium ions which are involved in a ternary complex model/Mg²⁺/ substrate (3). The occurence of this complex plays an important role in the stereoselective transfer of the hydrogen from the reagent to the substrate.

In our laboratory we have developped a series of chiral models possessing chiral aminoalcohol derivatives at the carbonyl at C-3 of the 1,4-dihydropyridine structure (4). In the ternary complex the rigidity of the chiral auxiliary is assured by the coordination of magnesium ions with the carbonyl group at C-3 and by the supplementary complexation caused by the alcoholic oxygen. By these means good e.e. is obtained with model bearing phenylalaninol at the carbonyl at C3.

In the past, in the thieno[2,3-b]pyridine series, we have shown that a model bearing a paranitrophenyl group instead of a phenyl group in phenylalaninol, leads to a good improvement of the e.e. (58 % to 75 %) (5). It was suggested that the rigidification of the chiral auxiliary in the ternary complex was enhanced by the establishment of a charge-transfer complex between the poor in electrons paranitrophenyl group and the rich in electrons thienodihydropyridine system.

A great improvement in the efficiency of these types of NADH models was carried out by studying

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reagents where the free rotating ability around the C3-C=O bond was suppressed. To attain this objective a cyclised reagent la was synthesized (6).

With this reagent an e.e. of about 90 % was obtained during the reduction of methyl benzoylformate at room temperature. The large enhancement of the e.e. is a result of the following two factors: i) the rigidity of the chiral auxiliary leads to a high stereodifferentiation of the two faces of the 1,4-dihydropyridine structure and ii) the readiness of the formation of the ternary complex.

It could be of interest to modulate the structure of the chiral auxiliary with a view to obtaining better e.e. and to disposing a larger field of performant reagents. With respect to this objective we synthesised and studied new cyclised models of type 1 derived of various chiral amino alcohols 9.

Scheme 1

The necessary chiral aminoalcohols were obtained in the following ways :

- norephedrine 9f is a commercial product.

- (S)-phenylalaninol 9a was obtained by reduction of phenylalanine methyl ester with NaBH₄ (5).

- (S)-paramethoxyphenylalaninol 9e, was obtained in a similar fashion after reduction of methylated tyrosine (5).

- (S)-aminoalcohols 9 b,c,d were also obtained by reduction of the corresponding (S) amino acid methyl esters. The latter were obtained by enantioselective enzymic hydrolysis (8) of racemic ethyl N-acetylamino acids using the following scheme (9) :

The optical purity of these aminoalcohols was established by GPC analysis of Mosher's derivatives of aminoesters **8b,c,d (10).** Compounds 8 and 9 were shown optically pure. It can be assumed that aminoalcohols $9b,c,d$ are also optically pure, the transformation $6 \rightarrow 9$ being non racemizing. All the available chiral aminoalcohols were involved in the synthesis of chiral NADH models 1 by using the previously described method (6b) (scheme 3).

Nota: it must be noted that an attempt to obtain a model with a (S)-paranitrophenylalaninol as chiral auxiliary was tried. But contrary to the behaviour in the thieno[2,3_b]pyridine series (5), we did not succeed in the reduction of the corresponding pyridinium salt with $Na₂S₂O₄$.

The reduction of methyl benzoyl formate at room temperature with these reagents leads **the results** shown in table 1.

Table 1^{$\frac{1}{2}$} **Reduction of methyl benzoylformate with various NADH models 1**

As can be seen the best e.e. is obtained with the β -naphtyl derivative $2a$. The enhancement in the efficiency due to the larger size of the naphtyl group, compared to the phenyl group in 1a is rather modest. By changing a phenyl group to a naphtyl group in other simpler NADH models the e.e. was improved from 20 % to 28 % (11).

The introduction dfla supplementary methylene group in the chain between the phenyl group and the

chiral carbon does not modify significantly the e.e. (model **lc** compared with model **la).** Despite the greater mobility of the hindering group, the stereodifferentiation of the two faces of the dihydropyridine is always well assured.

The results obtained with models **Id** and **le** are surprising. It could be assumed that the electron withdrawing CF_3 group would favour the formation of a charge transfer complex as was previously proposed for the paranitrophenyl group in the thienopyridine series (5). On the other hand the electron donating Me0 group would disfavour this charge transfer complex. As can be seen, the results do not confirm this hypothesis. We believe that, in the thienopyridine series, the charge transfer complex was the result of a strong interaction between the paranitrophenyl substituent and the π excessive thiophene ring.

The last substituent is derived from norephedrine. An examination of molecular models suggests that the methyle and phenyle groups could exert an additional influence on the stereodifferentiation of the two faces of the dihydropyridine. It appears that the behaviour of norephedrine is the same as phenylalaninol. However, norephedrine is a cheap, readily available commercial reagent and on this point alone, it can be claimed that model **If** is a readily available, highly enantioselective chiral NADH model.

EXPERIMENTAL

(S)-phenylalaninol 9a, (S>paramethoxyphenylalaninol 9e were obtained by previously described methods (5). Norephedrine **9f** was purchased from Aldrich.

I) *Synthesir of chiral amino* acids *by enzymic methoak*

I) Diethyl arylalkylacetamidomalonic acids : 2b,c,d (9). In a flask flushed with argon was introduced 100 ml of anhydrous ethanol and 2.3 g (0.1 mole) of sodium. After reaction and cooling to room temperature, 21.7 g (0.1 mol) of diethyl acetamidomalonate was added followed by, 10 minutes later, 0.11 mol of arylalkyl halogenated derivative $Ar(CH_2)_n$ Br. The mixture was heated to reflux for 15 hours. After cooling, derivatives 2 were isolated by filtration.

2b (n = 1, Ar = 2-naphtyl) yield 94 %. IR: v 1740 cm⁻¹; 1640 cm⁻¹. ¹H NMR (CDCl₂): 7.7 - 7.2 (m, 7H) ; 6.5 (s, 1H) ; 4.3 (q, 4H); 3.8 (s, 2H) ; 2.0 (s, 3H) ; 1,3 (t, 6H).

2c (n = 2, Ar = Ph) yield 60 %. IR: v 1740 cm⁻¹; 1640 cm⁻¹. ¹H NMR (CDCl₃): 7.2 (s, 5H); 6.9 (s, 1H); 4.2 (q, 4H); 2.6 (m, 4H); 2.0 (s, 3H); 1.2 (t, 6H).

2d (n = 1, Ar = p-CF₃Ph) yield 94 %. IR: 1740 cm⁻¹; 1650 cm⁻¹, ¹H NMR (CDCl₃): 7.5 (d, 2H); 7.1 (d, 2H); 6.6 (s, 1H); 4.3 (q, 4H); 3.7 (s, 2H); 2.0 (s, 3H); 1.3 (t, 6H).

2) *Ethyl arylakylacetamidomonomalonates : 3b,c,d.* The diester 2 (0.1 mol) was dissolved in 200 ml of ethanol. To this solution was added 75 ml of 3.9 M NaOH and the mixture was stirred at room temperature for 40 minutes. After acidification to pH=2, the ethanol was evaporated and the remaining solid was washed with water and dried.

3.5 (s, 2H); 1.9 (s, 3H); 1.1 (t, 3H). 3b yield 88 %. IR: v (C=O): 1770-1740-1610 cm⁻¹. ¹H NMR (DMSO-d₆): 8.0-7.0 (m, 9H); 4.1

3c yield 85 %. IR: v (C=O) : 1770-1740-1610 cm⁻¹. ¹H NMR (DMSO-d₆): 8.1 (s, 1H); 7.2 (s, 5H); 4.0 (q, 2H); 2.4 (s, 4H); 1.9 (s, 3H); 1.1 (t, 3H).

3d yield 85 %. IR: v (C=O) : 1770-1740-1620 cm⁻¹. ¹H NMR: (DMSO-d₆): 7.6 (d, 2H); 7.2 (d, 2H); 6.9 (s, 1H); 4.1 (q, 2H); 3.5 (s, 2H); 1.9 (s, 3H); 1.1 (t, 3H).

3) *Ethyl 2-arylalkyl-2 acetamidocarboxylates: 4b,c,d.* A solution of the monoester 3 (0.1 mol) in 400 ml of dioxane, was heated to reflux for 1 hour. After evaporation of the solvent, the residual oil was taken up in

4a yield 81 %. m.p.= 127°C. IR: v (C=O) 1750-1650 cm⁻¹. ¹H NMR (CDCl₃): 7.8-7.1 (m, 7H); 6.1 (d, 1H); 4.9 (m, 1H); 4.1 (q, 2H); 3.3 (d, 2H); 1.9 (s, 3H); 1.2 (t, 3H). Anal. Calcd for C₁₇H₁₉NO₃ (285.35): C, 71.56; H, 6.71; N, 4.91. Found: C, 71.65; H, 6.92; N, 4.75.

4b yield 78 %. m.p.= 72°C. IR: v (C=O) : 1750-1650 cm⁻¹. ¹H NMR (CDCl₃): 7.2 (m, 5H); 6.1 (d, 1H); 4.7 (m, 1H); 4.2 (q, 2H); 2.7 (m, 2H); 2.2 (m, 2H); 2.0 (s, 3H); 1.2 (t, 3H). Anal. Calcd for $C_{14}H_{19}N_0$ (249.32): C, 67.45; H, 7.68; N, 5.62. Found: C, 67.37; H 7.75; N, 5.79.

4c yield 80 %. m.p.= 112°C. IR: v (C=O): 1750-1650 cm⁻¹. ¹H NMR (CDCl₃): 7.6 (d, 2H); 7.2 (d, 2H); 6.2 (d, 1H); 4.9 (m, 1H); 4.2 (q, 2H); 3.2 (d, 2H); 2.0 (4, 3H); 1.2 (t, 3H). Anal. Calcd for $C_{14}H_{16}F_3N$ (303.30): C, 55.44; H, 5.32; N, 4.62. Found: C, 55.68; H, 5.32; N, 4.52.

4) *Enzymic resolution of racemic 4. Synthesis of (S)-2-arylakyl-2-aceiamidocarboxylic acids 6 and (R)-ethyl-2-arylalkyl-2 acetamidocarboxylic acids : 5.* Substrate 4 (0.04 mol) was dissolved in 60 ml of acetonitrile, 60 ml of water and 10 ml of a 0.01 M KCI solution. The mixture was vigorously stirred and 0.2 g of NaHCO₃ was added followed by 0.02 g of protease (subtilisin type VIII Sigma). The pH was maintained at 7.8 by use of an automatic burette delivering 10 μ fractions of 1M NaOH. The mixture was stirred at room temperature until there was no more NaOH used up (about 5 hours). After evaporation of the solvent the ester (R) -5 was separated by filtration. The aqueous phase was extracted with ethyl acetate, then acidified (pH = 1) with 6M HCI and the (S)-acid 6 was filtered and dried.

6b yield 48 %. m.p.= 208°C. IR: v (C=O): 1710-1630 cm⁻¹. ¹H NMR (DMSO-d₆): 8.2 (d, 1H); 7.8 (m, 3H); 7.7 (s, 1H); 7.4 (m, 3H); 4.5 (m, 1H); 3.3 (m, 2H); 1.8 (s, 3H). Anal. Calcd for $C_{15}H_{15}NO₃$ (257.30): C, 70.02; H, 5.88; N, 5.44. Found: C, 70.76; H, 6.03; N, 5.48.

6c yield 48 %. m.p.= 183°C. IR: v (C=O) : 1700-1630 cm⁻¹. ¹H NMR (DMSO-d₆): 8.2 (d, 1H); 7.2 (s, 5H); 4.1 (m, 1H); 2.5 (m, 2H); 1.9 (m, 5H). Anal. Calcd for C₁₂H₁₅NO₃ (221.26): C, 65.14; H, 6.83; N, 6.33. Found: C, 65.04; H, 6.72; N, 6.20.

6d yield 47 %. m.p. = 186°C. IR: v (C=O) : 1740-1610 cm⁻¹. ¹H NMR (DMSO-d₆): 8.2 (d, 1H); 7.6 (d, 2H); 7.3 (d, 2H); 4.4 (m, 1H); 3.0 (m, 2H); 1.8 (s, 3H). Anal. Calcd for C₁₂H₁₂F₃NO₃ (275.24): C, 52.36; H, 4.40; N, 5.09. Found: C, 52.24; H, 4.25; N, 4.88.

5) *Hydrolysis of compounds 6 : synthesis of(S)- amino acids 7. The* arylalkyl acetamidocarboxylic acid 6 (0.015 mol) was heated to reflux in 100 ml 6M HCI for 4 hours. After cooling, the water was evaporated to dryness and the amino acids were obtained in the form of hydrochloride.

6b, HCl : yield 90 %. m.p. > 250°C. IR: v (C=O) : 1730 cm⁻¹. ¹H NMR (DMSO-d₆): 8.6 (m, 3H); 7.9-7.4 (m, 7H); 4.2 (m, 1H); 3.3 (d, 2H). Anal. Calcd for $C_{13}H_{14}CINO_2$ (251.71): C, 62.03; H, 5.61; N, 5.57. Found: C, 61.63; H, 5.80; N, 5.46.

6c, HCl : yield 95 % m.p. > 250°C. IR: v (C=O) : 1730 cm v ¹. ¹H NMR (DMSO-d₆): 8.6 (m, 3H); 7.2 (s, 5H); 3.8 (m, 1H); 2.6 (m, 2H); 2.1 (m, 2H). Anal. Calcd for $C_{10}H_{14}CINO_2$ (251.68): C, 55.69; H, 6.54; N, 6.49. Found: C, 56.42; H: 6.63; N, 6.57.

6d, HCl : yield 90'% m.p.= 246 °C. IR: v (C=O) : 1730 cm⁻¹. ¹H NMR (DMSO-d₆): 8.6 (m, 3H); 7.7 (d, 2H); 7.5 (d, 2H); 4.2 (μ i, 1H); 3.3 (d, 2H). Anal. Calcd for C₁₀H₁₁ClF₃NO₂ (269.66): C, 44.54; H, 4.11; N, 5.19. Found: C, 44.56; IH, 4.04; N, 5.09.

II) Synthesis of chit&

I) (S)-methyl aminoesters 8. To the above chloride of amino acid 7 (0.015 mol) suspended in 15 ml of methanol was added 3 rtil of redistilled thionyl chloride (0.041 mol). The solution was heated to reflux for 2 hours. After eliminati the form of an I volatile products, the remaining solid was dried and the eater 8 was obtained in hydrocl

8b, HCl yield 95%. m.p.= 195°C. IR: v (C=O) : 1750 cm⁻¹. ¹H NMR (DMSO-d₆): 8.8 (m, 3H); 7.85

(m, 4H); 7.45 (m, 3H); 4.35 (m, 1H); 3.7 (s, 3H); 3.3 (t, 2 H). Anal. Calcd for $C_{14}H_{16}CINO_2$ (265.75): C, 63.28; H, 6.07; N, 5.27. Found: C, 63.47; H, 6.03; N, 5.17.

8c, HCI yield 94 % m.p.= 194 °C. IR: v (C=O) : 1750 cm⁻¹. ¹H NMR (DMSO-d₆) : 8.8 (m, 3H); 7.2 (s, SH); 3.8 (m, 1H); 3.6 (s, 3H); 2.6 (m, 2H); 2.1 (m, 2H). Anal. Calcd for $C_{11}H_{16}CINO_2$ (229.71): C, 57.51; H, 7.02; N, 6.10. Found: C, 57.45; H, 6.54; N, 5.89.

8d, HCl yield 94 %. m.p.= 200°C. IR: v (C=O) : 1740 cm⁻¹. ¹H NMR (DMSO-d₆): 8.8 (m, 3H); 7.7 (d, 2H); 7.5 (d, 2H); 4.3 (t, 1H); 3.7 (s, 3H); 3.3 (t, 2H). Anal. Calcd for $C_{11}H_{13}CIF_3NO_2$ (283.68): C, 46.57; H, 4.62; N, 4.94. Found: C, 46.49; H, 4.62; N, 4.89.

2) *Determination of optical purity of aminoesters Ib,c,d* Racemic esters 4b, 4c, and 4d have been prepared by the method precedently described without enzymic resolution. In this way racemic aminoacids **6b,c,d were** obtained and they were transformed into racemic esters **Ib,c,d.** For the determination of the optical purity, the Mosher's derivatization method was used (10).

Into a test tube was introduced 5 mg of racemic aminoester. The tube was flushed with argon and 300 μ l of anhydrous pyridine was added followed by 50 pl of Mosher's acid chloride. The mixture was stirred for 10 minutes at room temperature. The excess Mosher's reagent was destroyed by adding 50 pl of methanol and the resulting mixture was analysed by GPC.

- chromatograph DELSI SI200

- injector "split" 25O'C

- detector FID 25O"C

- column DB1 capillary $30x0.25x0.25$ (m x mm x μ m)

- vector gas : hydrogen (1 bar)

- oven temperature : 200° C for c,d ; 240° C for b.
- retention times (approximately, in minutes) :

Under the same conditions it was observed that the (S)-aminoesters obtained after enzymic resolution were optically pure.

3) *Reduction of (S)-aminoesters 8b,c,d. Synthesis of (S)-aminoalcohols 9,b,c,d.* To a solution of 1.75 g of NaBH₄ (0.046 mol) in 25 ml of a mixture water/ethanol (1/1) was added a solution of aminoester 8 and 25 ml of the same solvent mixture. The resulting mixture was heated to reflux for 4.5 hours, then the ethanol was evaporated. The aqueous phase was saturated with sodium chloride and extracted with ethyl acetate. After drying and evaporation of the solvent a pale yellow oil was obtained which soon crystallised.

9b yield 78 % m.p.= 112'C. 'H NMR (CDCI,): 7.8 (m, 3H); 7.6 (s, 1H); 7.4 (m, 2H); 7.3 (m, 1H); 3.7 (dd, **1H);** 3.4 (dd, 1H); 3.2 (m, 1H); 3.0 (dd, IH) ; 2.7 (dd, 1H). Anal. Calcd for C,,H,,NO (201.27): C, 77.58; H, 7.51; N, 6.96. Found: C, 77.71; H, 7.29; N, 6.67.

9c yield 72 % m.p.= 76°C. ¹H NMR (CDCl₃): 7.2 (m, 5H); 3.6 (dd, 1H); 3.3 (dd, 1H); 2.8 (m, 1H); 2.6 (m, 2H); 1.6 (m, 2H).

9d yield 75 96 m.p.= 74"C. 'H NMR (CDC13): 7.6 (d, 2H); 7.3 (d, 2H); 3.6 (dd, 1H); 3.4 (dd, 1H); 3.1 (m, 1H); 2.9 (dd, 1H); 2.6 (dd, 1H). Anal. Calcd for C₁₀H₁₂F₃NO (219.22): C, 54.79; H, 5.52; N, 6.39.
Found C, 54.67; H, 5.39; N, 6.02. *III) Synthesis of models I*

I) S-oxo-5,6,7,8-titrahydro-Z,6-naphtyridines: 12. A solution of 1.28 g (0.010 mol) of 2-ethynylnicotinonitrile and 0.011 mol of chiral aminoalcohol 9 in 35 ml of anhydrous THF was heated to reflux for 24 hours. The solvent was evaporated and the crude enamine **10 was** dissolved in JO ml of methanol. To the solution was added 1.3 g of NaBH₃CN and 1.4 g of anhydrous ZnCl₂. The mixture was stirred at room temperature. One hour later 0.65 g of NaBH₃CN and 0.7 g of ZnCl₂ were added and stirring was maintained for 1 hour. After cooling, a solution of 0.1 M NaOH was added and the mixture was extracted with CH₂Cl₂. After drying followed by evaporation of the solvent, crude compound 11 was obtained and not further purified. This product was dissolved in 20 ml of a mixture ethanol/water (95/5) and stirred for 48 hours. After evaporation of the solvents, the resulting solid was purified by column chromatography (silica, eluent CH₂Cl₂/EtOH : 90/10). (The yields are given with respect to 2-ethynylnicotinonitrile).

12s yield 56 % m.p.= 125'C (6b)

12b $\bf H$ ield 45 % m.p.= 122°C. IR: v (C=O) : 1620 cm⁻¹. ¹H NMR (CDCl₃): 8.5 (dd, 1H); 8.25 (dd, 1H); 7.7 (m, 4); 7.4 (m, 3H); 7.25 (dd, 1H); 4.85 (m, 1H); 3.9 (m, 2H); 3.5 (m, ZH); 3.2 (m, 3H); 2.9 (m, 2H). Anal. Calcd for C₂₁H₂₀N₂O₂ (332.40): C, 75.88; H, 6.06; N, 8.43. Found: C, 76.03; H, 5.89; N, 8.53.

12c yield 46 %. IR: u (C=O) : **1630 cm- '. 'H NMR (CDCl): 8.6 (dd, 1H); 8.3** (dd, 1H); 7.2 (m, 6H); 4.8 (m, 1H); 3.8-3.6 (m, 5H); 3.2 (m, 2H); 2.7 (m, 2H); 1.9 (m, 2H). Anal. Calcd for $C_{18}H_{20}N_2O_2$ (296.37): C, 72.95; H, 6.80; N, 9.45. Found: C, 72.76; H, 6.72; N, 9.29.

12d yield 39 % m.p.= 136°C. IR: v (C=O) : 1640 cm⁻¹. ¹H NMR (CDCl₃): 8.55 (dd, 1H); 8.25 (dd, 1H); 7.55 (d, 2H); 7.35 (d, 2H); 7.25 (dd, 1H); 4.80 (m, 1H) ; 3.90 (m, 2H); 3.50 (m, 2H); 3.10 (d, 2H); 2.95 (m, 3H). Anal. Calcd for $C_{18}H_{17}F_3NO_2$ (350.35): C, 61.71; H, 6.42; N, 7.99. Found: C, 61.28; H, 5.02; N 8.19.

12e yield 71 % m.p.= 104°C. JR: v (C=O) : 1625 cm- l. 'H NMR (CDCI,): 8.52 (d, 1H); 8.23 (d, 1H); 7.22 (dd, 1H); 7.13 (d, 2H); 6.77 (d, 2H); 4.78 (m, IH); 3.73 (s, 3H); 3.65 (m, 2H); 3.48 (t, 2H); 2.92 (m, 4H). Anal. Calcd for C₁₈H₂₀N₂O₃ (312.37): C, 69.21; H, 6.45; N, 8.97. Found: C, 69.05; H, 6.51; N, 8.93.

12f yield 40 %. IR; v (C=O): 1630 cm⁻¹. ¹H NMR (CDCl₃): 8.3 (dd, 1H); 8.05 (dd, 1H); 7.2 (m, 6H); 4.07 (m, 3H); 3.4 (m, 2H); 2.75 (m, 2H); 1.1 (d, 3H). Anal. Calcd for $C_{17}H_{18}N_2O_2$ (282.34): C, 72.32; H, H, 6.35. N, 9.83.

2) Quaternization of compounds 12: Synthesis of pyridinium salts 13. Compound 12 (0.004 mol) was dissolved in 5 ml of acetonitrile. To the solution, 5 ml of methyl iodide was added and the mixture was heated to reflux for 12 hours. After cooling, $2/3$ of the volatile compounds were evaporated and diethyl ether was added. The pyridinium sailt which precipitated was filtered. The yield was nearly quantitative.

13a (6b)

: v (C=O) : 1670 cm⁻¹. ¹H NMR (DMSO-d₆): 9.0 (d, 1H); 8.7 (d, 1H); 8.0 (t, 1H); 7.8 (m, 4H); 7.4 (m, 3H); 5.0 (m, 2H); 4.2 (s, 3H); 3.7 (m, 4H); 3.3-3.1 (m, 4H). Anal. Calcd for $C_{22}H_{23}IN_2C$ (474.34): C, 55.71; H, 4.89; N, 5.90. Found: C, 55.11; H, 4.79; N, 5.73.

 $=$ O) : 1670 cm⁻¹. ¹H NMR (DMSO-d₆): 9.10 (d, 1H); 8.90 (d, 1H); 8.10 (t, 1H); 7.20 (m, 5H); 4.60 (m, 1H); 4.25 (s, 3H); 3.70-3.40 (m, 6H); 2.55 (m, 2H); 1.80 **(m,** 2H). Anal. Calcd for $C_{19}H_{23}$ $\mathbb{N}_{2}O_{2}$ (438.30): C, 52.07; H, 5.29; N, 6.39. Found: C, 52.19; H, 5.24; N, 6.20.

 $=$ O) : 1670 cm⁻¹. ¹H NMR (DMSO-d₆): 9.05 (d, 1H); 8.75 (d, 1H); 8.0 (t, (m. 2H): 4.25 (s, 3H); 3.65 (m. 4H); 3.3 (m, 2H); 3.0 (d, 2H). Anal. Calcd for $C_{19}H_{20}F_3IN_2O_2$ (492.28): C, 46.36; H, 4.09; N, 5.69. Found: C, 46.51; H, 4.31; N, 5.63.

: v (C=O) : 1670 cm⁻¹. ¹H NMR (DMSO-d₆) : 9.06 (d, 1H); 8.78 (d, 1H); 8.02 (t, 2H); 4.92 (t, 1H); 4.82 (m, 1H); 4.25 (s, 3H); 3.67 (s, 3H); 3.60 (m, 4H); 3.25 (m, $2H$); 2.82 (m, 2H). Anal. Calcd for $C_{10}H_{23}N_2O_3$ (454.3): C, 50.23; H, 5.10; N 6.16. Found: C, 49.74; H, 4.84; N, 5.94.

13f m.p.= 202°C. IR: v (C=O) : 1660 cm⁻¹. ¹H NMR (DMSO-d₆): 9.07 (d, 1H); 8.78 (d, 1H); 8.03 (t, 1H); 7.25 (m, 5H); 5.62 (d, 1H); 4.71 (m, 2H); 4.21 (s, 3H); 3.65 (m, 2H); 3.25 (m, 2H); 1.10 (d, 3H). Anal. Calcd for C₁₈H₂₁IN₂O₂ (424.23): C, 50.96; H, 4.99; N, 6.60. Found: C, 50.75; H, 4.97; N, 6.61.

3) Reduction of pyridinium salts 13 : synthesis of models 1. The pyridinium salt (0.001 mol) was dissolved at 40°C in 5 ml of water which have been degsssed with argon. After dissolution the mixture maintained under argon was returned to room temperature and 0.53 g of $Na₂CO₃$ was added. A solution of 0.70 g of $Na_2S_2O_4$ in 3 ml of degassed water was then added with vigorous stirring. The precipitate was filtered and washed with degassed water, then dried. These compounds were not purified further.

la (6b)

lb yield 60 %. ¹H NMR (CDCl₃): 7.8 (m, 3H); 7.6 (s, 1H); 7.4 (m, 3H); 5.6 (d, 1H); 4.7 (q, 1H); 4.2 (m, 2H); 3.8 (m, 2H): 3.3-3.0 (m. 6H); 2.8 (s, 3H); 2.1 (m, 2H).

lc yield 86 %. 'H NMR (CDCls): 7.25 (m, SH); 5.7 (d, 1H); 4.75 (q, 1H); 4.35 (m, 1H); 3.7 (m, 2H); 3.3 (m, 2H); 3.15 (m, 2H); 2.95 (s, 3H); 2.65 (m, 2H); 2.4 (m, 2H); 1.9 (m, 2H).

1d yield 90 %. ¹H NMR (CDCl₃): 7.55 (d, 2H); 7.35 (d, 2H); 5.65 (d, 1H); 4.75 (q, 1H); 4.1 (m, 2H); 3.75 (m, 2H); 3.2-3.0 (m, 6H); 2.85 (s, 3H); 2.2 (m, 2H).

le yield 88 %. 'H NMR (CDCls): 7.13 (d, 2H); 6.82 (d, 2H); 5.65 (d, 1H); 4.76 (q, 1H); 4.15 (m, 2H); 3.79 (m. 5H); 3.2-2.9 (m, 6H); 2.86 (s, 3H); 2.15 (t, 2H).

1f yield 88 %. ¹H NMR (CDCl₃): 7.29 (m, 5H); 5.67 (d, 1H); 4.90 (s, 1H); 4.80 (q, 1H); 4.40 (m, 1H); $3.15-2.75$ (m, 7H); 2.25 (t, 2H); 1.18 (d, 3H).

4) *Reduction of methyl phenylglyoxyfate with models 1.* For the procedure and the determination of e.e. see ref (7).

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